HIV Testing

Conventional laboratory tests and some new developments *Molecular Diagnosis*

NSW State Reference Laboratory for HIV Centre for Applied Medical Research St Vincent's Hospital Sydney





This presentation

- Refresher on conventional diagnostic testing for HIV
- New developments with 'Point of Care' rapid tests
- Molecular testing for HIV diagnosis and management
- Some applications

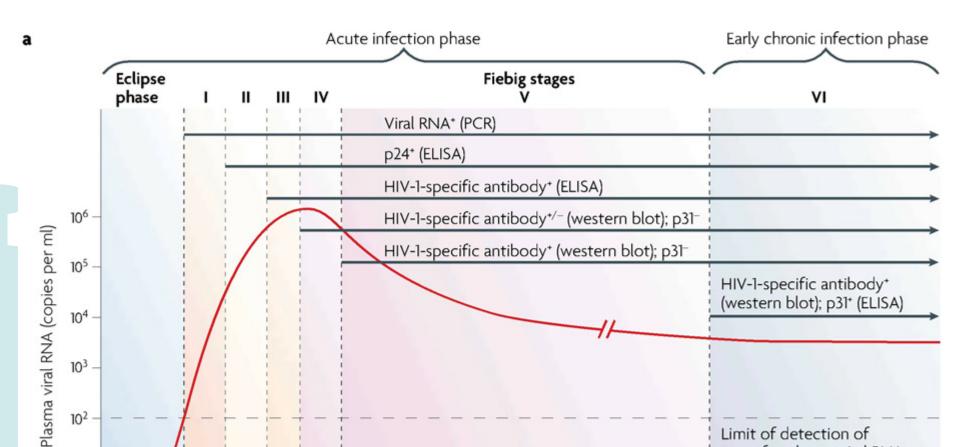




History of HIV testing in Australia

- HIV testing introduced 1985 in Australia and first to have safe blood supply, with blood bank screening also introduced in 1985
- ●1987 HIV Testing was described in the 1st National HIV/AIDS Strategy
- Free, confidential testing backbone of HIV prevention
- Seroconversion illness characterised in Australia & targeted lab testing became the norm
- There is emerging concern that HIV testing rates may be dropping







Limit of detection of assay for plasma viral RNA

100

50

Days following HIV-1 transmission

40

10²

10¹

10

20

30

Table 1. HIV testing assays and their "window periods."

| HIV test | Assay method | "Window period" estimates, weeks ^a | "Window period" reduction, days ^b |
|-----------------------|--|--|---|
| First-generation EIA | Viral particles used to bind patient HIV Ab, detected by marker conjugated to anti-human Ab | ~6 | |
| Second-generation EIA | Same as first-generation EIA except uses purified HIV Ag or re- combinant virus | ~4–6 | 10 |
| Third-generation EIA | "Antigen sandwich": synthetic peptide used to bind patient HIV Ab followed by marker conjugated to additional HIV Ag; able to detect IgM | ~3–4 | 6 |
| Fourth-generation EIA | Uses third-generation EIA methodology plus monoclonal Ab to p24 Ag to detect patient p24 Ag | ~2 | 5 |
| Pooled HIV NAT | First combines multiple individual samples into one common pool, then uses PCR or other amplification techniques to detect patient viral nucleic acids | <1–2 | 3 |
| Individual HIV NAT | As above, except that samples are tested individually rather than diluted by pooling | <1-2 | 3 |

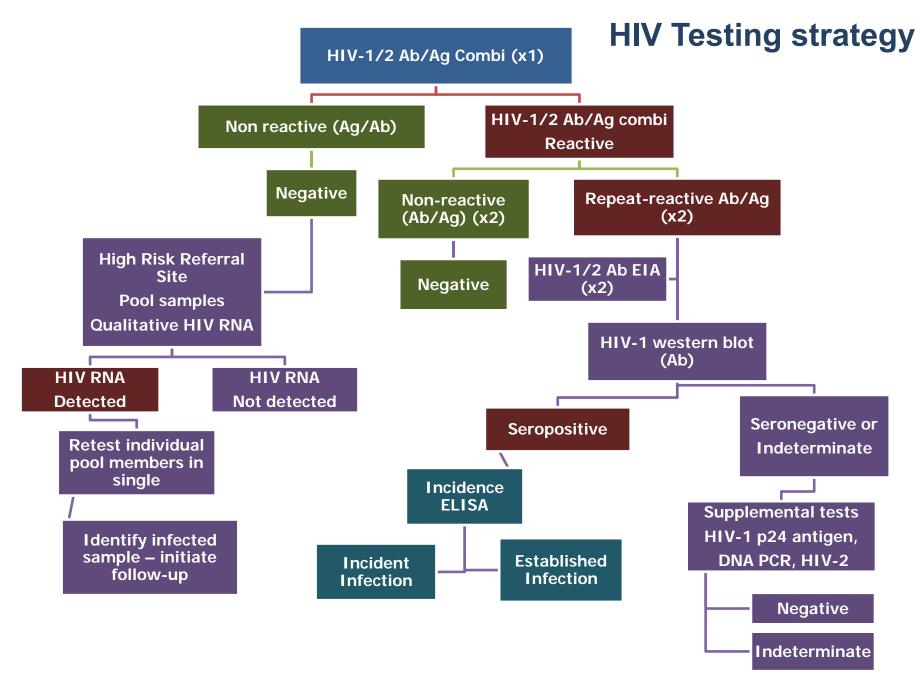
NOTE. Data are from [5, 6, 16, 19]. Ab, antibody; Ag, Antigen; HIV NAT, HIV nucleic acid testing.

460 • CID 2007:44 (1 February) • HIV/AIDS



^a "Window periods" listed are averages. For example, although second generation EIAs will detect HIV infection in nearly all individuals within 6 weeks of HIV acquisition, in a study of occupationally exposed health care workers, 5% of subjects did not have seroconversion until at least 6 months following the exposure [20].

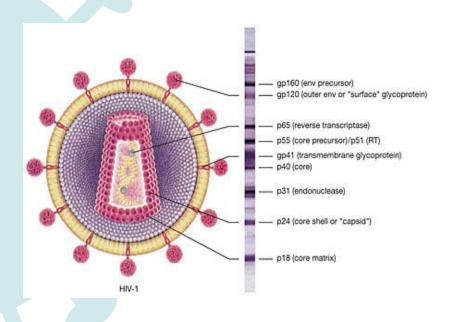
^b Compared with an immediate less sensitive assay, the "window period" for pooled HIV NAT is, on average, 3 days shorter than the "window period" for a fourth-generation EIA.

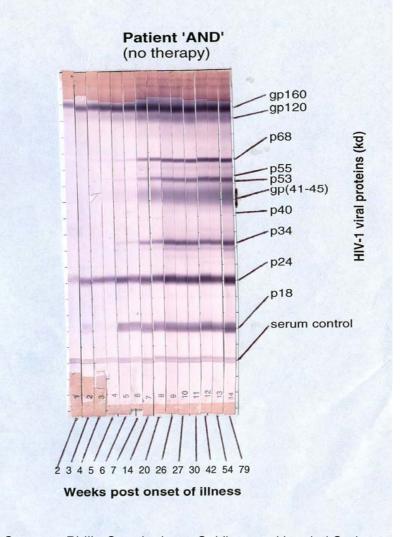


Serology of primary HIV-1 infection

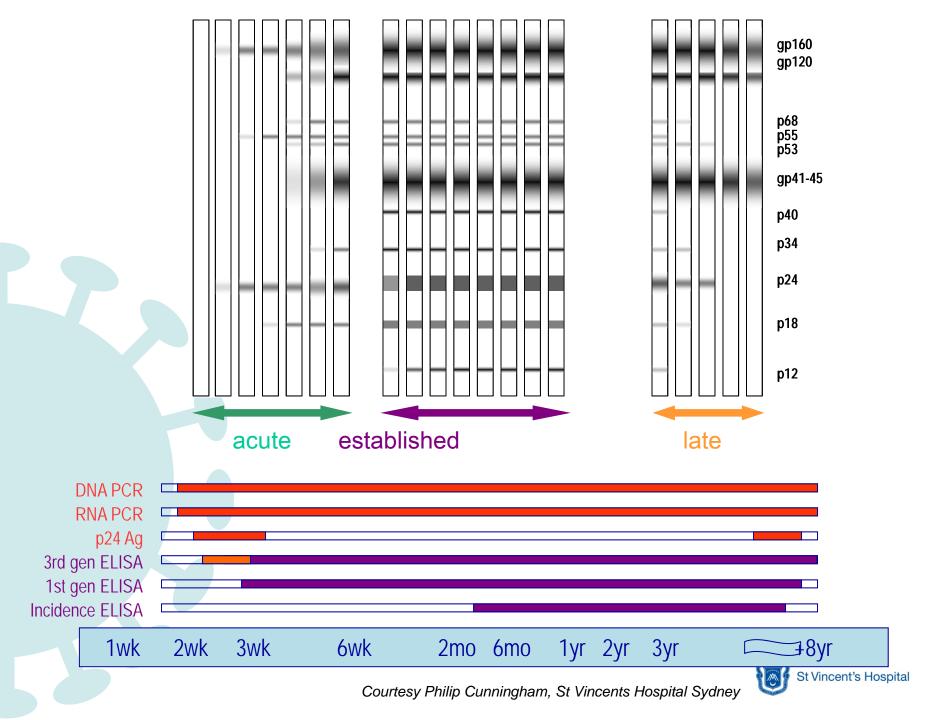
(Western Blot)

- Antibody tests may be negative or low level (indeterminate)
- Antibodies to viral proteins are incrementally detected – increasing in intensity over time (diagnostic)





Courtesy Philip Cunningham, St Vincents Hospital Sydney



Western blot criteria

NEGATIVE: No reactivity to any viral specific bands

POSITIVE: Reactivity to at least one (1) envelope

glycoprotein (gp41-45, gp120 or

gp160), and at least three (3) viral

specific bands of the HIV-1 gag or pol

gene product series



1 2 3 4 5 6 7 8 9 0 1 2 3 4 5 6 7 gp160 gp120 p68 p55 p51 gp41 p40 p34 p24 p18

Western blot reactivity interpretation

| 1 | Indeterminate group 1 – likely biological false positive | p55 |
|----|--|--|
| 2 | Positive – advanced HIV infection, late stage (seroreversion) | gp160, gp120, p68, p55, p51, gp41 |
| 3 | Positive - established HIV infection | gp160, gp120, p68, p55, p51, gp41, p40, p34, p24, p18 |
| 4 | Positive – recent HIV infection – seroconversion from group 4 indeterminate to positive profile | gp160, gp120, p68, p55, p51, p24 |
| 5 | Indeterminate group 4 – advanced HIV infection, late stage (seroreversion) | gp160, gp120, p68, gp41 |
| 6 | Positive – recent seroconversion in acute HIV infection | gp160, p55, p24, p18 |
| 7 | Indeterminate group 4 – acute HIV infection | gp160, p55, p24 |
| 8 | Indeterminate group 4 – acute HIV infection | gp160, p55, p24 |
| 9 | Indeterminate group 3 — either acute infection or non specific reactivity, require supplemental test results or HIV DNA nucleic acid | p24 |
| 10 | Indeterminate group 2 – probable non-specific reactivity | p18 |
| 11 | Indeterminate group 3 - either acute infection or non specific reactivity, require supplemental test results or HIV DNA nucleic acid | p24 |
| 12 | Indeterminate group 2 – probable non-specific reactivity | p18 |
| 13 | Positive – advanced HIV infection , late stage(seroreversion) | gp160, gp120, p68, p55, p51, gp41, p24 |
| 14 | Positive – advanced HIV infection , late stage (seroreversion) | gp160, gp120, p68, p55, p51, gp41, p18 |
| 15 | Positive – advanced HIV infection, late stage (seroreversion) | gp160, p68, p51, gp41, p34 |
| 16 | Negative (negative control) | no viral bands |
| 17 | Positive – established HIV infection (positive control) | gp160, gp120, p68, p55, p51, gp41, p40, p34, p24, p18 |
| | | St Vincent's Hospital |

Primary HIV Infection: Why are we so interested?

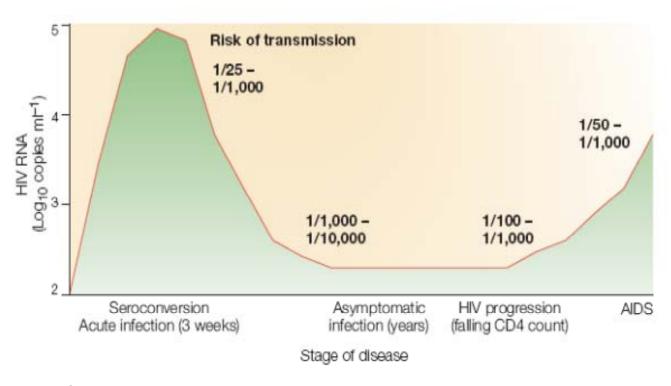
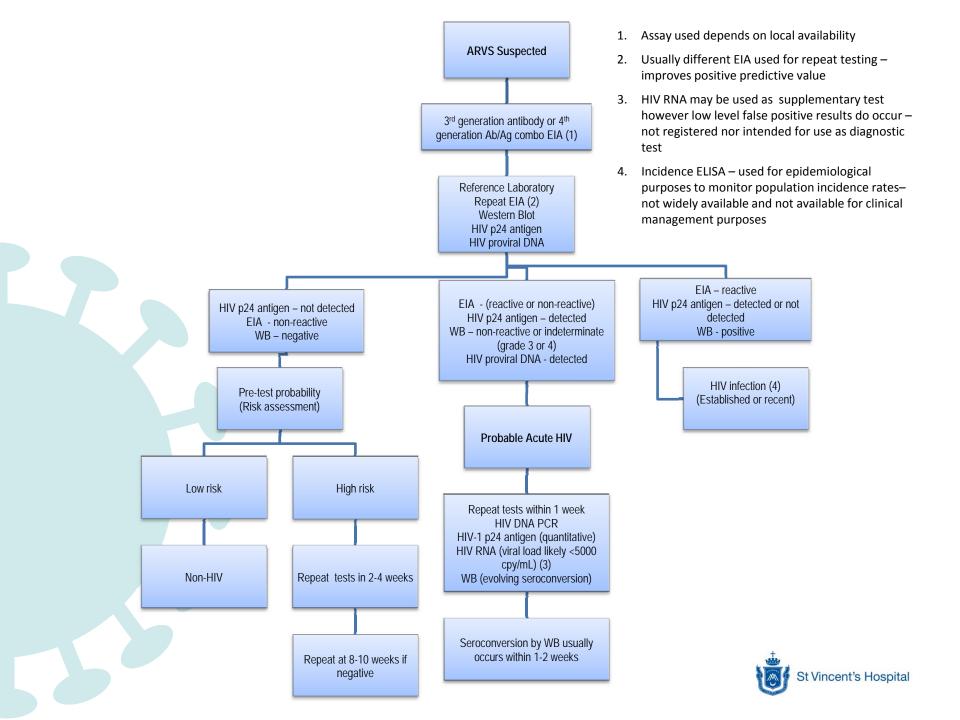


Figure 1 | The changing viral load during the different stages of disease and the effects of viral load on the probability of sexual transmission of HIV.





HIV Testing Policy

■The first testing policy was introduced in 1998 it was written by a committee and endorsed by the Commonwealth

Subsequently updated in 2006, but only issued as a PDF document and available via websites







Point of Care Testing

- Short incubation tests
- ■Test which can be taken to the person being tested, rather than the sample having to be transported to another location
- Can be done using:
 - Capillary blood dropped onto a sample well which is soaked with a reagent or into a chamber to which reagent is added
 - Oral fluid swabbed from the gum line





Lot number Patient Identification Name of Test HINTER Test ID number Read Control Result Here Read Patient Result Here Add Sample Here

Determine[™] HIV-1/2

Whole Blood Procedure

(Refer to package inserts for assay procedures) (Refer to the other side for Serum/Plasma procedure)

Remove tests



Note: Removal of the test units should start from the right side of the test card to preserve the lot number which appears on the left side of the card.

Remove cover

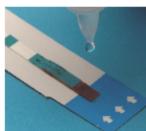


Add sample



Add sample (50µl) to sample pad (finger stick or venipuncture)

Add chase buffer



Add one drop of chase buffer

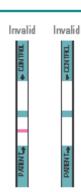






Control Patient





1 minute

determine detect.... diagnose... determine...

www.determinetest.com

enquiry@determinetest.com

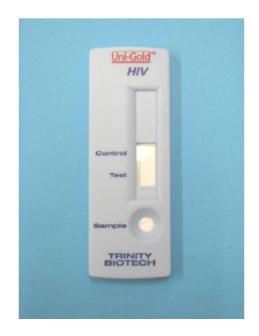


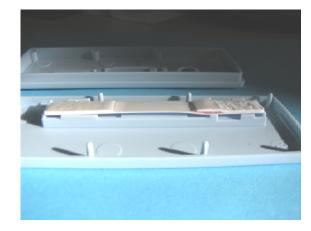
Immunochromatography

- ↓ IgG Ab
- Y α-HIV Ab
- ▲ HIV-Ag
- [▲] α −hu-lgG
- label

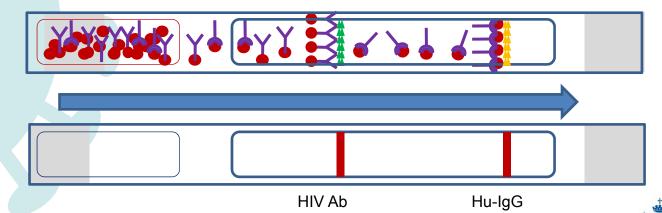








St Vincent's Hospital





Advance Awareness™







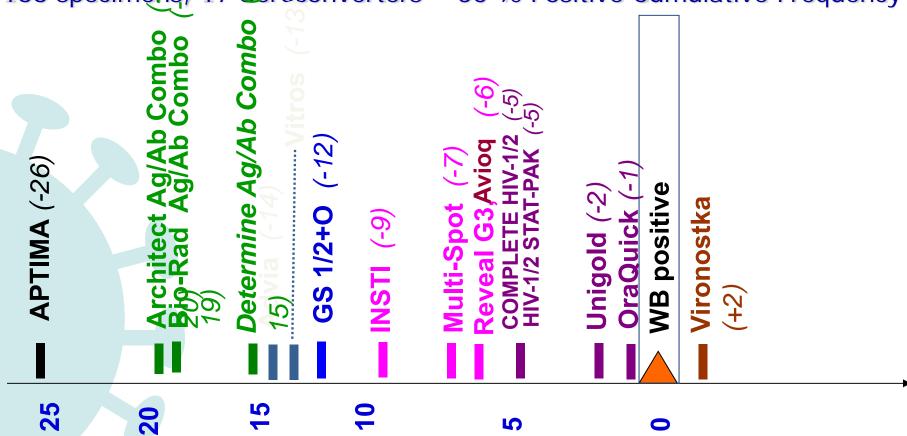






Sequence of Test Positivity Relative to WB

166 specimens, 17 Seroconverters - 50 % Positive Cumulative Frequency



Days before WB positive

Modified from Masciotra et al, J Clin Virol 2011 and Owen et al, J Clin Micro 2008





Demand for & on point of care tests

Increase testing rates



Increase engagement with care



Increase # people on treatment



Will failure to identify acute infection cause a problem





HIV Testing Direct Detection of Virus

Virus isolation – PBMC co-culture

- Insensitive
- Requires containment laboratory (PC3)
- ~ 3-6 weeks culture
- Detect with p24 antigen or RT activity in culture supernate

HIV-1 p24 antigen

- Simple serology serum no additional sample required
- Sensitivity about 30 copies HIV RNA (10pg)
- Now incorporated in 4th generation screening tests
- Limited use during acute and ? Advanced or late infection

Nucleic acid detection - (NAT)

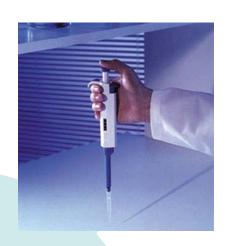
- Qualitative tests more sensitive and specific blood/tissue screening
- Most products available are quantitive viral load tests intended use is NOT diagnostic
- Circulating cell free RNA or integrated cellular DNA ??
- Dedicated samples required
- Laboratory facilities



NAT tests for HIV

| DNA | RNA | RNA |
|--|--|--|
| Qualitative | Quantitative (viral load) | Qualitative |
| <10 copies | <50 copies | <10 copies |
| Highly specific | Specificity issues at low range | Highly specific |
| HIV supplemental diagnostic test | HIV monitoring test | Blood and tissue donor screening |
| Resolve indeterminate serology Acute infection diagnosis (pre-seroconversion) Early infant diagnosis | Treatment responsePrognostic markerClinical decision point | • Detection of HIV in pooled or single blood/tissue donors |
| Integrated and unintegrated cellular DNA | Extracellular RNA (free virion) | Extracellular RNA (free virion) |
| Whole blood Dried Blood Spot (DBS) | Plasma DBS | Plasma Cadaveric blood |

Pre amplification - extraction





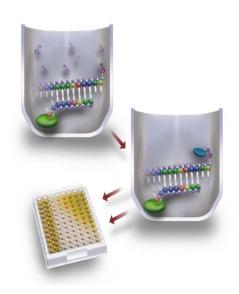


- Volumetric errors amplified
- •Tedious manual, repetitive
- Specimen integrity



Post amplification & detection Endpoint detection







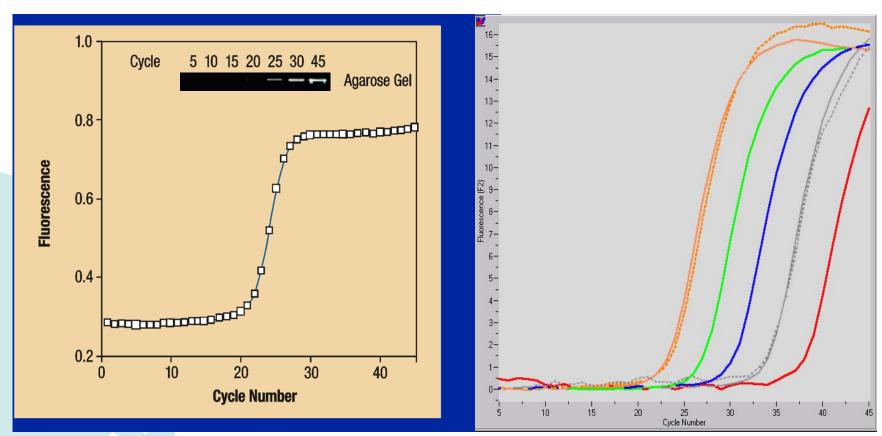
- Volumetric error
- •Fragment size vs. probe hybridisation
- Time to result
- Automation calibration issues
- Result calculations







Monitoring in Real time









Real time PCR























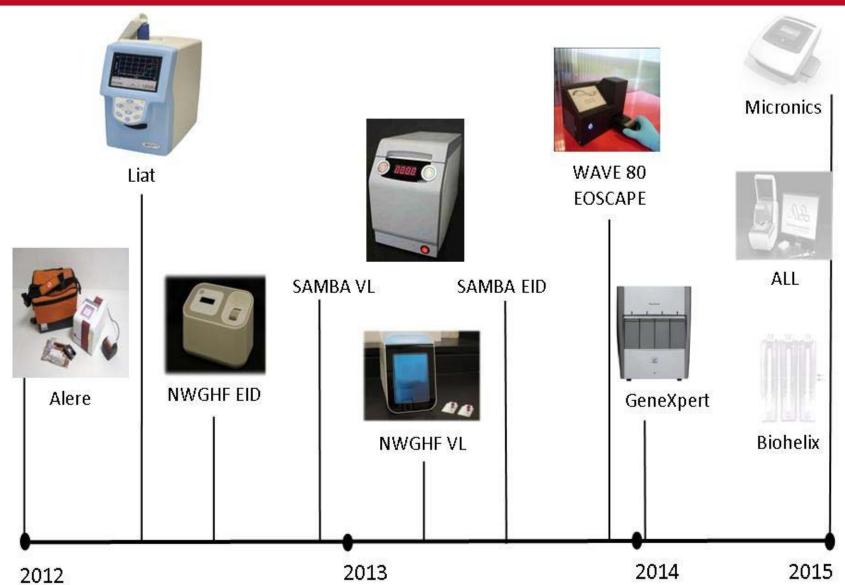
SIEMENS







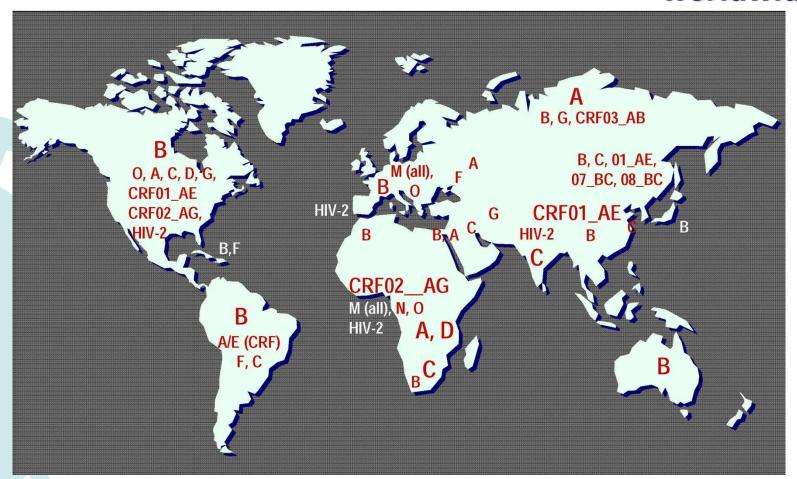
Point-of-Care Viral Load and EID Technologies in the Pipeline*



^{*}Estimated; timeline and sequence may change.

UNITAID is hosted and administered by the World Health Organization © 2011 World Health Organization (Acting as the host organization for the Secretariat of UNITAID)

Geographic distribution of HIV Subtype B: represents ~10% of HIV-1 infections worldwide





Increasing Spread of HIV Diversity





Continual redistribution of HIV variants

Put science on your side.

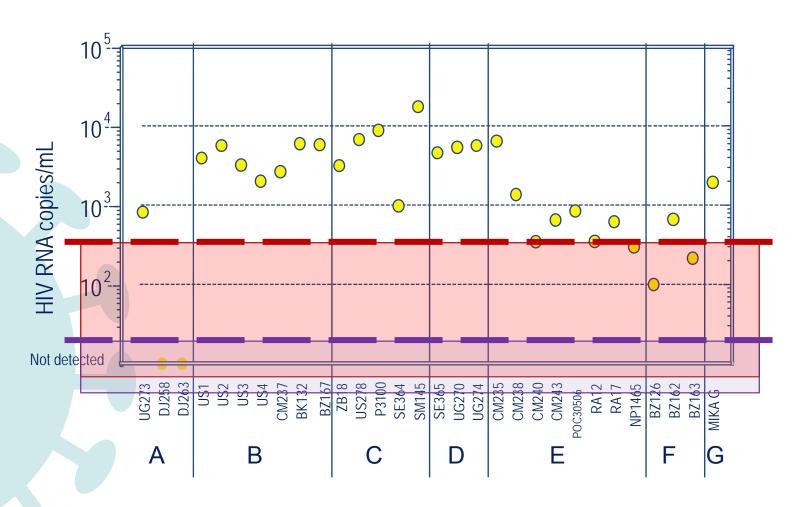
Date: 12-01-09

Company Confidential © 200X Abbott 27



Courtesy: Dr John Hackett

Qualitative vs quantitative assays



Isolate and Subtype



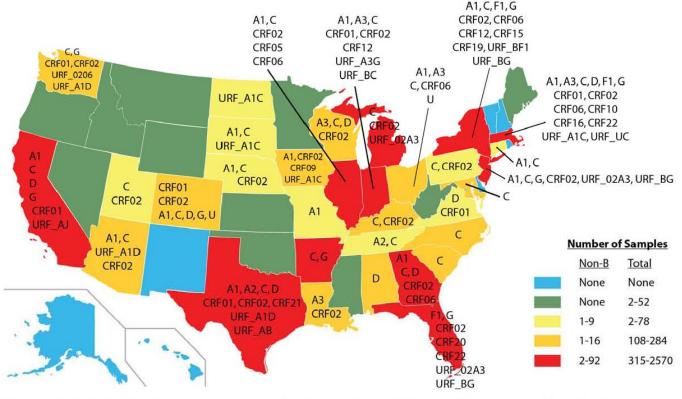
HIV subtype distribution in USA

• n = 12,650; 43 states

• 392 non-B strains (3.1%; 32 states)

• 5 subtypes; 13 CRFs; 24 URFs

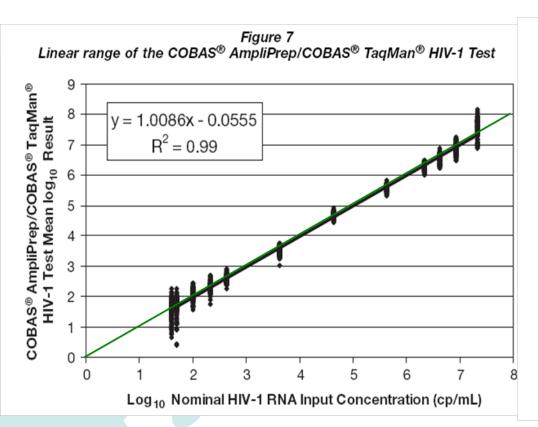
Pyne *et al* 16th CROI (2009); 292 Data thru Sept 09

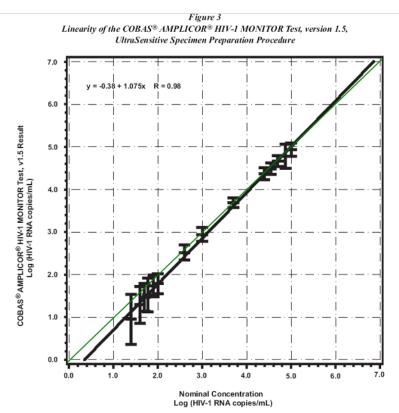


<u>Geographic distribution of non-B subtypes.</u> Samples were received from 43 states, 32 of which included non-B subtype samples. The non-B subtypes are listed for each state. The colors indicate the total number of samples received and the number of non-B samples received from each state.



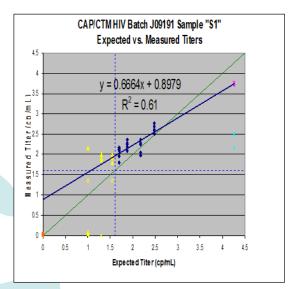
Real time versus end-point detection COBAS Taqman vs COBAS Amplicor HIV MONITOR Accuracy at lower limit of detection

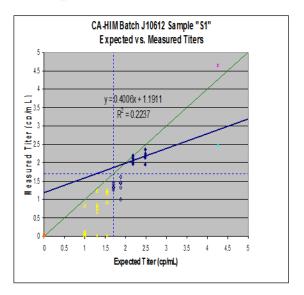


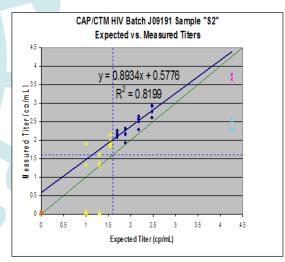


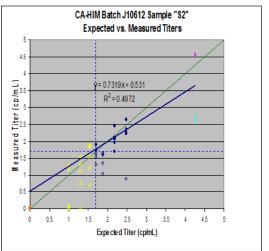


Real time versus end-point detection COBAS Taqman vs COBAS Amplicor HIV MONITOR Accuracy at lower limit of detection









Source: Roche Molecular



Early infant diagnosis – Qualitative HIV DNA testing





Diagnostic Testing in Infants

- In infants <18 mos, use virologic assays that directly detect HIV: HIV DNA PCR or HIV RNA (AII)
 - Persistent maternal antibody give false positive antibody results
- Virologic testing of HIV-exposed infant (AII)
 - 14-21 days of age
 - 1-2 months
 - 4-6 months
- Virologic testing at birth should be considered for infants at high risk of infection (BIII)
- HIV antibody test
 - 12-18 months to document seroreversion in HIV-uninfected infants
 (BIII)
 - Diagnostic test for children ≥18 months (AII)





Criteria for HIV Diagnosis

- 2 positive HIV virologic tests on separate blood samples (regardless of age) (AII)
- Positive HIV antibody test with confirmatory Western blot (or IFA) at age ≥18 months (AII)





Development of a DBS collection SOP









Venous whole blood collected into EDTA tubes may be used to prepare DBS samples within 24 hours from collection @ 4°C.





For early infant diagnosis a heel prick sample is collected as shown. Encourage blood flow by warming the site with soft cloth moistened with warm water up to 41°C, or by rubbing for 3-5 minutes



Wash hands with soap and water and wear clean gloves for each new patient.



Have card ready and labeled with 2x patient identifiers (name + DOB) PLUS date of collection. DO NOT TOUCH the collection circles





Disinfect the site with alcohol wipe dry with cotton, puncture site, allow drop of blood to form. Touch drop onto card avoid layering. Completely fill 4 circles



Blood should saturate each circle. Collect four circles per patient. Follow universal safety precautions



For venous blood Clearly label card with identification number Pipette volume required to uniformly saturate entire circle (usually 50-75 µl)



Collection of Dried Blood Spot (DBS)

Early infant HIV diagnosis, HIV serology HIV drug resistance testing and HIV viral load



Dry DBS at Room temperature Fully air dry (≥4 hours, or overnight)Use a drying rack if possible

Do not touch or smear spots Do not dry in direct sunlight Do not heat, stack, or allow DBS to touch each other or other surfaces while drying





Suitable DBS should be saturated through card 10



poorly collected DBS. These samples are Unsuitable for testing



Package dried DBS in zip-locked plastic bag, add desicant and humidity indicators. Seal plastic bad and label outside of bag

with permanent marker







Keep packaged DBS (in sealable plastic bags) cool and dry until transported to reference laboratory within 2 weeks.

Do not leave in vehicle, as sun and heat will deteriorate DBS.





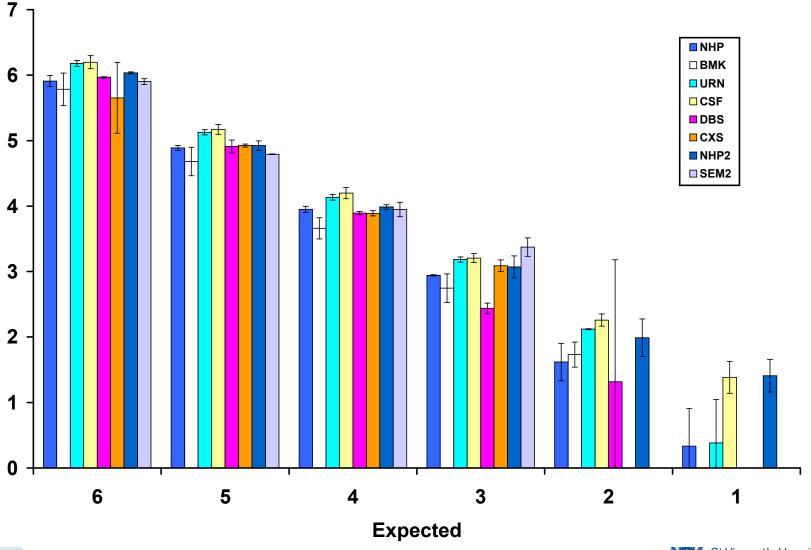
- Insert bundled DBS into rip-resistant envelope.
- Include appropriate documentation.
- Insert both into envelope and seal for shipment.



Send samples to 14 laboratory. Note poorly collected samples will affect the quality of the results. DBS must be clearly labeled, saturate 4 circles and completely dried and remain dry.



DBS for HIV viral load?



A Loftis, R Kshatriya, K McCall-Culbreath, S Fiscus and J Nelson. IAS 2009

