Point of Care Tests for respiratory viruses: impact on clinical outcomes of patients

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Virology Research Laboratory (VRL), UNSW and POWH













Outline

- 1. Importance of respiratory viral infections
- 2. POCT tests available
- 3. Studies using POCT what are the real-world outcomes?
- 4. New developments in POCT
- 5. Summary







Importance of respiratory viral infections

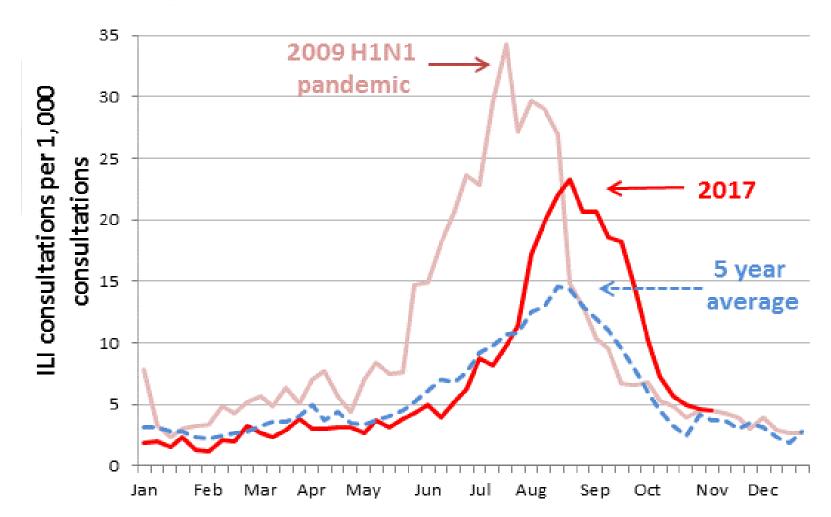








Importance of respiratory viral infections



Source: ASPREN and VIDRL







Respiratory viral infections cause exacerbations

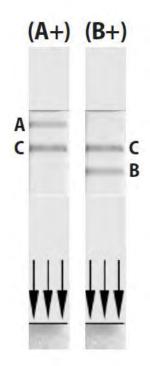
- Chronic respiratory disease: asthma, cystic fibrosis (CF) and chronic obstructive pulmonary disease (COPD)
- Worsened during exacerbations
 - → influenza and rhinovirus
- Flu vaccine → 33% effective in 2017 (Sullivan et al., 2017)
 → 11% in people ≥65 y.o.
- Antivirals available but time critical



Types of POCT for respiratory viruses

Lateral flow POCT

Quidel Quickvue











Types of POCT for respiratory viruses

Lateral flow POCT

Positives:

- Easy-to-use
- Results in ≤15 minutes
- Excellent specificity
- Compatible with multiple transport options
- Long shelf-life

Negatives:

- ↓ sensitivity
- Influenza B sensitivity lower than influenza A
- Influenza A virus subtype ?
- Limited multiplex







BinaxNOW

Study	Study Population	Sample type	Sensitivity	Specificity
Hassan <i>et al.,</i> 2014 J Clin Micro	Pediatric	200 frozen NP swabs & washes	Flu A- 72.8% Flu B- 70.8%	Flu A- 100% Flu B- 100%
Cho <i>et al.,</i> 2013 J Virol Methods	Adult and pediatric	253 frozen NP swabs	Flu A- 71% Flu B- 37.2%	Flu A-100% Flu B- 100%
DiMaio <i>et al.,</i> 2012 J Virol Methods	Adult and pediatric	200 frozen NP samples	Flu A-62.2% Flu B- 54.5%	Flu A-100% Flu B- 100%
Booth <i>et al.,</i> 2006 J Med Virol	Adult and pediatric	224 frozen NPA & N/T swabs	Flu A-80% Flu B- 47%	Flu A- 99% Flu B-100%







Types of POCT for respiratory viruses

- Lab in a capsule POCT
- All-in-one machines





GenePOC and PIE Up to 12 targets



- Cdiff
- VRE
- Staph. Aureus
- Multi-Drug Resistance CRE
- GBS
- Pharyngeal strep
- Flu/RSV
- HIV-HBV-HCV
- Enteric Panels
- Respiratory Panels
- Mycobacterium
- CT/GC/TV/MG
- Vaginitis Panel
- HSV







Types of POCT for respiratory viruses

Lab in a capsule POCT

Positives:

- Excellent specificity and sensitivity
- Traceability
- Influenza A subtyping
- Other viruses

Negatives:

- Expensive
- Require specialised equipment







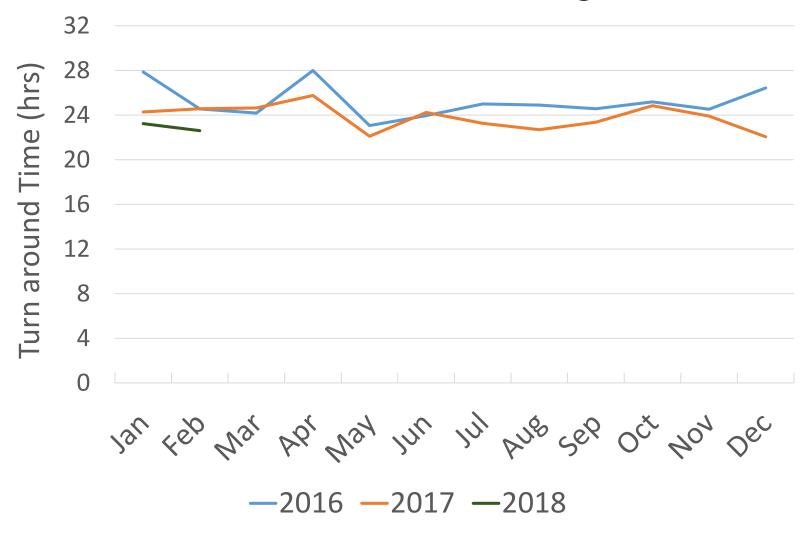
Cepheid GeneXpert







POWH PCR Testing

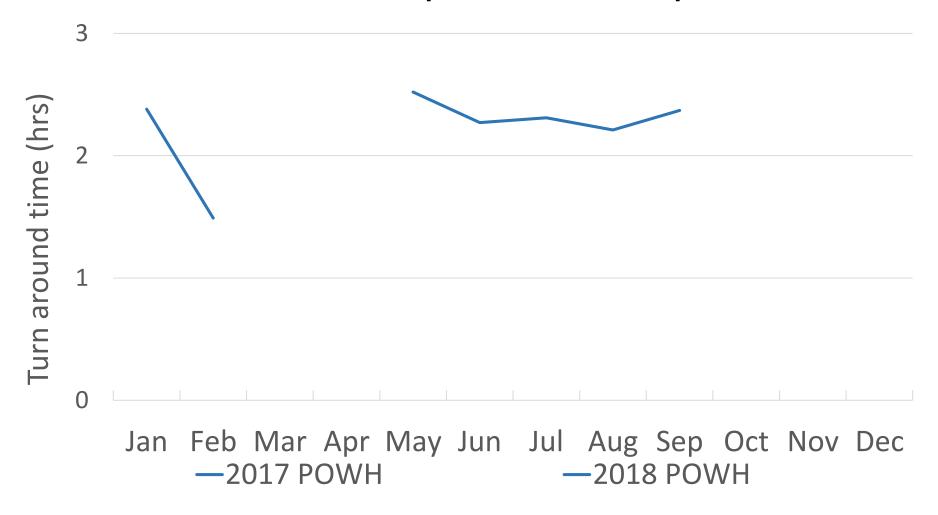








POWH Cepheid GeneXpert









Potential benefits of POCT

- ↑ prescription of antivirals
- prescription of antibiotics
- ↓ length of hospital stay
- ↓ exacerbations

Does this happen in practice?







- Systematic review:
 - i) antiviral prescription
 - ii) antibiotic prescription
 - iii) patient length of stay in the ED
- Comprehensive search of all primary research papers available that meet search criteria

Egilmezer et al., 2018, Reviews in Medical Virology, provisionally accepted







Clearly defined eligibility criteria & methodology

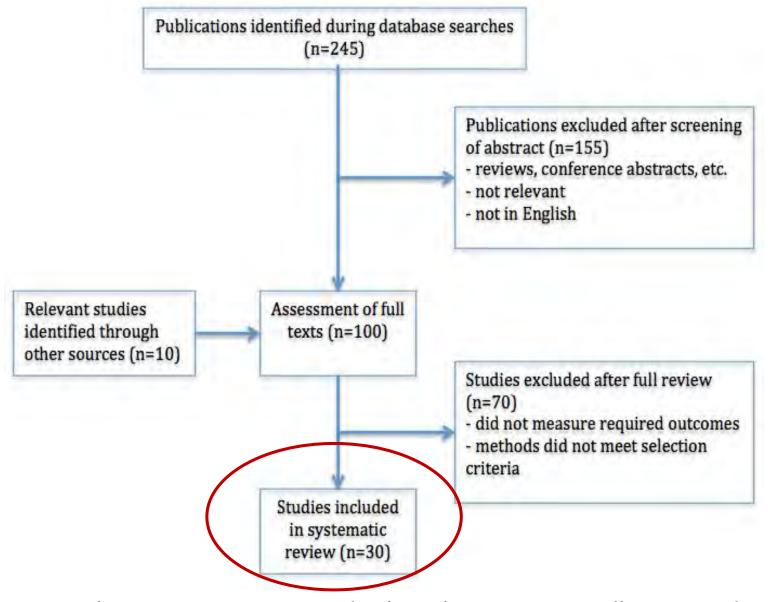
Medline & Embase

• "influenza, point-of-care test, antivirals, antibiotics, length of stay"









Egilmezer et al., 2018, Reviews in Medical Virology, provisionally accepted







Antiviral prescription: 14 studies

↑ 12/14 (86%)

Antibiotic prescription: 26 studies

↓ 20 / 26 (77%)

Egilmezer et al., 2018, Reviews in Medical Virology, provisionally accepted







Time spent in ED: 9 studies

5/9 (55.6%) \

More research needed







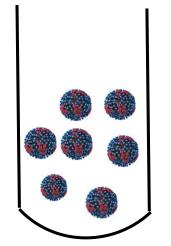
Developing New POCT

- Aim: develop methods to rapidly detect pathogen nucleic acid using "everyday" devices
- Low-middle income countries

Target antigen binding, triggering release of fluorescent molecules.

→ Readouts using commonly available













Developing New POCT

- Prof Justin Gooding
- Dr Padma Bakthavathsalam
- NSW Smart Sensing Network (NSSN)







- → Reduce overuse of antibiotics
- → antibiotic resistance







What are the features of new POCT devices?

- Simple
 - clear instructions, straightforward read-out
- Cheap
- No specialised machinery
- Robust reagents and consumables
- Concordance with established laboratory methods sensitive and specific
- Low sample volume
- Safe (devices, reagents, disposal)
- Traceability









Developing New POCT

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Anal Chem. 2015 August 4; 87(15): 7872–7879. doi:10.1021/acs.analchem.5b01594.

Paper-Based RNA Extraction, *in Situ* Isothermal Amplification, and Lateral Flow Detection for Low-Cost, Rapid Diagnosis of Influenza A (H1N1) from Clinical Specimens

Natalia M. Rodriguez[†], Jacqueline C. Linnes[†], Andy Fan[†], Courtney K. Ellenson[†], Nira R. Pollock[‡], and Catherine M. Klapperich^{†,*}

[†]Department of Biomedical Engineering, Boston University, Boston, Massachusetts 02215, United States

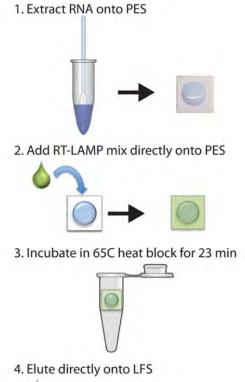
[‡]Division of Infectious Diseases, Beth Israel Deaconess Medical Center, Harvard Medical School, Boston, Massachusetts 02115, United States

Abstract

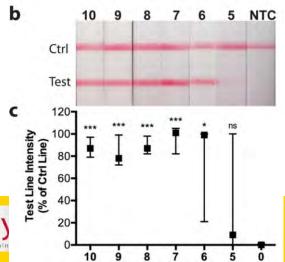
The 2009 Influenza A (H1N1) pandemic disproportionately affected the developing world and high-lighted the key inadequacies of traditional diagnostic methods that make them unsuitable for use in resource-limited settings, from expensive equipment and infrastructure requirements to unacceptably long turnaround times. While rapid immunoassay diagnostic tests were much less costly and more context-appropriate, they suffered from drastically low sensitivities and high false

PES- poly(ether sulfone) paper matrix









Summary

- Improved clinical outcomes:
 - ↑ prescription of antivirals
 - ↓ prescription of antibiotics
 - ? stay in ED
- Improved prognostic data
- Potential use of new POCT for control of outbreaks of known and emerging respiratory viruses







Thank you

Virology Research Lab

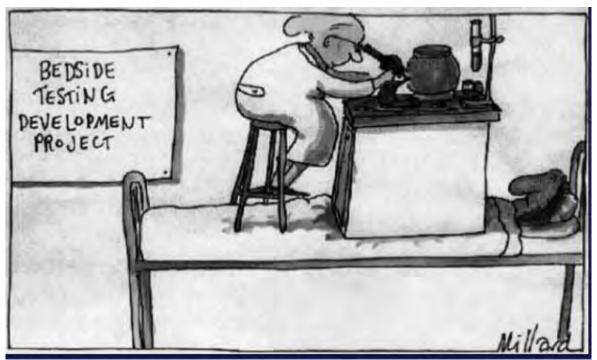
- Gregory Walker
- Ece Egilmezer

UNSW Chemistry

- Prof Justin Gooding
- Dr Padma
 Bakthavathsalam
- Danielle

Integrated Sciences

Dylan Warby









Questions

- Does anyone currently use POCT?
- What system do you use?
- What would be the ideal POCT if we could design from scratch?







Thank you

Virology Research Lab

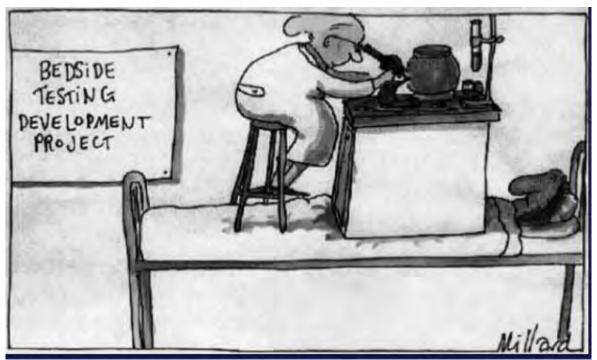
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Randomised controlled trial and health economic evaluation of the impact of diagnostic testing for influenza, respiratory syncytial virus and Streptococcus pneumoniae infection on the management of acute admissions in the elderly and high-risk 18- to 64-year olds

Nicholson K G, Abrams K R, Batham S, Medina M J, Warren F C, Barer M, Bermingham A, Clark T W, Latimer N, Fraser M, Perera N, Rajakumar K & Zambon M.

Detailed Author information

Health Technology Assessment Volume: 18, Issue: 36, Published in May 2014

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https://doi.org/10.3310/hta18360 📑

Citation: Nicholson K. Abrams K. Batham S. Medina M. Warren F. Barer M. et al. Randomised controlled trial and health economic evaluation of the impact of diagnostic testing for influenza, respiratory syncytial virus and Streptococcus pneumoniae infection on the management of acute admissions in the elderly and high-risk 18- to 64-year olds. Health Technol Assess 2014;18(36)





Toolkit

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Paper-Based RNA Extraction, *in Situ* Isothermal Amplification, and Lateral Flow Detection for Low-Cost, Rapid Diagnosis of Influenza A (H1N1) from Clinical Specimens

Natalia M. Rodriguez[†], Jacqueline C. Linnes[†], Andy Fan[†], Courtney K. Ellenson[†], Nira R. Pollock[‡], and Catherine M. Klapperich[†],*

[†]Department of Biomedical Engineering, Boston University, Boston, Massachusetts 02215, United States

[‡]Division of Infectious Diseases, Beth Israel Deaconess Medical Center, Harvard Medical School, Boston, Massachusetts 02115, United States

Abstract

The 2009 Influenza A (H1N1) pandemic disproportionately affected the developing world and high-lighted the key inadequacies of traditional diagnostic methods that make them unsuitable for use in resource-limited settings, from expensive equipment and infrastructure requirements to unacceptably long turnaround times. While rapid immunoassay diagnostic tests were much less costly and more context-appropriate, they suffered from drastically low sensitivities and high false negative rates. An accurate, sensitive, and specific molecular diagnostic that is also rapid, low-cost, and independent of laboratory infrastructure is needed for effective point-of-care detection and epidemiological control in these developing regions. We developed a paper-based assay that allows for the extraction and purification of RNA directly from human clinical nasopharyngeal specimens through a poly(ether sulfone) paper matrix, H1N1-specific *in situ* isothermal amplification directly within the same paper matrix, and immediate visual detection on lateral flow strips. The complete sample-to-answer assay can be performed at the point-of-care in just 45 min, without the need for expensive equipment or laboratory infrastructure, and it has a clinically relevant viral load detection limit of 10⁶ copies/mL, offering a 10-fold improvement over current



GenePOC

Instrument

Characteristics



- Compact / Portable
- Easy to use (2 min hands on time)
- Fast (70m turnaround time)
- Innovative (multiplexing up to 12 targets)
- Flexible (up to eight samples and 8 different assays per run)

PIE

Menu of Assays

- Cdiff
- VRE
- Staph. Aureus
- Multi-Drug Resistance CRE
- GBS
- Pharyngeal strep
- Flu/RSV
- HIV-HBV-HCV
- Enteric Panels
- Respiratory Panels
- Mycobacterium
- CT/GC/TV/MG
- Vaginitis Panel
- HSV







Quidel Quickvue

Study	Study Population	Sample type	Sensitivity	Specificity
Koul <i>et al.,</i> 2015 Indian J Med Microbiol.	Adult and pediatric	600 N/T swabs	Flu A- 22.7% Flu B- 23.6%	Flu A-100% Flu B-100%
Lucas <i>et al.,</i> 2011 Clin Infect Dis	Adult and pediatric	1538 nasal wash	Flu A- 15% H1N1 ₀₉ -20% Flu B- 31%	Flu A- 99% H1N1 ₀₉ - 99% Flu B- 99%
Velasco et al., 2010 J Clin Virol.	Adult and pediatric	360 nasal swabs	Flu A H1N1 ₀₉ - 63%	Flu A H1N1 ₀₉ - 96%
Company Reported		Nasal swab	FluA – 94% Flu B – 70%	FluA – 90% Flu B – 97%





